Instructions for Use

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Human Double-Stranded DNA IgM (dsDNA IgM) ELISA Kit

Catalog No: abx159293

Size: 96T

Range: Qualitative

Sensitivity: Qualitative

Storage: Store at 2-8°C for 6 months.

Application: The quantitative detection of dsDNA IgM in Human serum.

Principle of the Assay: This kit is based on enzyme-linked immuno-sorbent assay technology. An antigen is pre-coated onto a 96-well plate. Controls and test samples are added to the wells and incubated. The HRP-conjugated reagent is then added, and the whole plate is incubated. Unbound conjugates are removed using wash buffer at each stage. TMB substrate is used to quantify the HRP enzymatic reaction. After TMB substrate is added, only wells that contain sufficient dsDNA IgM will produce a blue coloured product, which then changes to yellow after adding the acidic stop solution. The intensity of the yellow colour is proportional to the dsDNA IgM amount bound on the plate. The Optical Density (OD) is measured spectrophotometrically at 450 nm in a microplate reader, from which the concentration of dsDNA IgM can be calculated.

Kit Components

- Pre-coated 96-Well Microplate: 12 x 8
- Wash Buffer (25X): 20 ml
- Positive Control: 800 µl
- Negative Control: 800 µl
- Sample Diluent Buffer: 20 ml
- Detection Reagent: 10 ml
- TMB Substrate A: 5 ml
- TMB Substrate B: 5 ml
- Stop Solution: 5 ml
- Plate Sealer: 4
- Hermetic Bag: 1

Materials Required But Not Provided

- 37°C incubator
- Multi and single channel pipettes and sterile pipette tips
- Squirt bottle or automated microplate washer
- 1.5 ml tubes
- Distilled water
- Absorbent filter papers
- 100 ml and 1 liter graduated cylinders
- Microplate reader (wavelength: 450 nm)
- ELISA Shaker

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Protocol

A. Sample Preparation

Analyse immediately or store samples at 2-8°C (within 24 hrs). For long term storage, aliquot and store at -20°C or -80°C. Avoid multiple freeze-thaw cycles.

- **Serum:** Samples should be collected into a serum separator tube. Coagulate the serum by leaving the tube undisturbed in a vertical position overnight at 4°C or at room temperature for 2 hrs. Centrifuge at approximately 1000 × g for 15 mins. If precipitate appears, centrifuge again. Assay immediately or aliquot and store at -20°C or -80°C. Avoid repeated freeze/thaw cycles. Avoid hemolytic samples.

Notes:

- **Analyse samples at a 101-fold (1/101) dilution (i.e. add 2 µl of sample to 200 µl of Sample Diluent Buffer).**
- Always use non-pyrogenic, endotoxin-free tubes for blood collection.
- Fresh samples, or recently obtained samples, are recommended to prevent protein degradation and denaturation that may lead to erroneous results.
- NaN_3 cannot be used as a test sample preservative, since it inhibits HRP.
- If a sample is not indicated in the manual's applications, a preliminary experiment to determine the suitability of the kit will be required.

B. Reagent Preparation

Wash Buffer: Dilute the concentrated Wash buffer 25-fold (1/25) with distilled water (i.e. add 20 ml of concentrated wash buffer into 480 ml of distilled water). If crystals have formed in the concentrated Wash Buffer, warm to room temperature and mix gently until the crystals have completely dissolved.

C. Assay Protocol

Prepare all standards, samples and reagents as directed above. Equilibrate the kit components and samples to room temperature prior to use. It is recommended to measure in duplicate, and to plot a standard curve for each test.

1. Set test sample, positive control, negative control, and blank (zero) wells on the pre-coated plate respectively, and record their positions. *Add the solution to the bottom of each well without touching the side walls. Pipette the standards and samples up and down to mix before adding to the wells. Avoid foaming or bubbles.*
2. Aliquot 100 µl of Positive Control into the positive control wells.
3. Aliquot 100 µl of Negative Control into the negative control wells.
4. Leave the blank (zero) well empty without any liquid.
5. Aliquot 100 µl of appropriately diluted sample (**101-fold**) into the test sample wells. Gently tap the plate to mix, or use a microplate shaker.
6. Cover the plate with a plate sealer and incubate for 30 mins at 37°C.
7. Remove the cover and discard the solution. Wash the plate 5 times with 1X Wash Buffer. *Fill each well completely with Wash buffer (350 µl) using a multi-channel Pipette or autowasher (1-2 mins soaking period is recommended). Complete removal of liquid at each step is essential for good performance. After the final wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean absorbent paper towels.*
8. Aliquot 100 µl of Detection Reagent to each well except the blank well. Seal the plate and incubate for 30 mins at 37°C.
9. Remove the cover, discard the solution and repeat the wash process as described above, 5 times.
10. Aliquot 50 µl of TMB Substrate A and 50 µl of TMB Substrate B into each well. Cover the plate with the plate sealer. Gently tap the plate to mix thoroughly. Incubate at 37°C for 10-20 minutes. The incubation time is for reference only, the optimal time should be determined by end user. Avoid exposure to light.
11. Aliquot 50 µl of Stop Solution into each well. It is important that the Stop Solution is mixed quickly and uniformly throughout the microplate to inactivate the enzyme completely.
12. Ensure that there are no fingerprints or water on the bottom of the plate, and that the fluid in the wells is free of bubbles. Measure the OD at 450 nm immediately.

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Data Analysis:

For the test to be valid, the Mean OD of Negative Control should be < 0.15, and the Mean OD of Positive Control should be > 0.6

CUT OFF value = Mean OD of Negative Control + 0.1 (if Negative Control < 0.1, calculate at 0.1; otherwise, calculate at the actual value)

If OD of Samples ≥ CUT OFF, the test samples are considered positive.
If OD of Samples < CUT OFF, the test samples are considered negative.

Precautions:

- Before using the kit, centrifuge the tubes to bring down the contents trapped in the lid.
- Do not leave the wells uncovered for extended periods between incubations. The addition of reagents for each step should not exceed 10 mins.
- Ensure that the plate is properly sealed or covered during the incubation steps, and that the time and temperature are controlled.
- Do not reuse pipette tips and tubes.
- Do not use expired components, or components from a different kit.
- The TMB substrate should be used under sterile conditions, and light exposure should be minimised. Unused substrate should be colorless, or a very light yellow in appearance. Do not discard any residual solution back into the vial.
- Please note that this kit is optimised for detection of native samples, rather than recombinant proteins or synthetic chemicals. We are unable to guarantee detection of recombinant proteins, as they may have different sequences or tertiary structures to the native protein.

Precision:

Intra-assay Precision (Precision within an assay): 3 samples of known concentration were tested 20 times on one plate.

Inter-assay Precision (Precision between assays): 3 samples of known concentration were tested on 20 assays.

CV (%) = (Standard Deviation / Mean) × 100

Intra-Assay: CV < 15%

Inter-Assay: CV < 15%

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