

Human Secretory Immunoglobulin A (sIgA) CLIA Kit

Catalog No.: abx197661

Size: 96T

Range: 62.5 pg/ml - 4000 pg/ml

Sensitivity: 37.5 pg/ml

Storage: Store the 96-well plate, Standards, HRP-conjugate reagent and Biotin-conjugated antibody at -20°C, and the rest of the kit components at 4°C for up to 6 months.

Application: For quantitative detection of sIgA in Human Serum, Plasma and other biological fluids.

Introduction: Immunoglobulin A (IgA) is an antibody that plays a crucial role in the immune function of mucous membranes. The amount of IgA produced in association with mucosal membranes is greater than all other types of antibody combined. In absolute terms, between three and five grams are secreted into the intestinal lumen each day. This represents up to 15% of total immunoglobulins produced throughout the body. IgA has two subclasses (IgA1 and IgA2) and can be produced as a monomeric as well as a dimeric form. The IgA dimeric form is the most prevalent and is also called secretory IgA (sIgA). IgA is a poor activator of the complement system, and opsonises only weakly.

Principle of the Assay

This kit is based on sandwich chemiluminescent immunoassay technology. Anti-sIgA antibody is pre-coated onto a 96-well plate. The standards and samples are added to the wells and incubated. Biotin-conjugated anti-sIgA and avidin conjugated to HRP are added to each microplate well and incubated. After the mixture of substrate A and B is added only wells that contain sIgA will produce chemiluminescence. The intensity of the emitted light is proportional to the amount of sIgA in the sample or standard.

Kit components

1. One pre-coated 96-well microplate (12 × 8 well strips)
2. Standard: 2 tubes
3. Sample/Standard Diluent Buffer: 20 ml
4. Biotin conjugated antibody (Dilution 1:100): 120 µl
5. Antibody diluent buffer: 12 ml
6. HRP Conjugate Reagent (Dilution 1:100): 120 µl
7. HRP diluent buffer: 12 ml
8. Substrate Reagent A: 5 ml
9. Substrate Reagent B: 5 ml
10. Wash buffer (25X): 30 ml
11. Plate Sealer: 5

Material Required But Not Provided

1. 37°C incubator
2. Luminometer capable of reading 96-well microplates
3. Multi and single channel pipettes and sterile pipette tips
4. Squirt bottle or automated microplate washer
5. 1.5 ml tubes to prepare standard/sample dilutions
6. Absorbent filter papers
7. 100 ml and 1 liter graduated cylinders

Protocol

A. Preparation of sample and reagents

1. Sample

Store samples to be assayed within 24 hours at 2-8°C. Alternatively, aliquot and store at -20°C or -80°C for long term. Avoid repeated freeze-thaw cycles.

- **Serum:** Samples should be collected into a serum separator tube. Coagulate the serum by leaving the tube undisturbed in a vertical position overnight at 4°C or at room temperature for up to 60 minutes. Centrifuge at approximately 1000 × g for 15 min. Analyze the serum immediately or aliquot and store at -20°C or -80°C.
- **Plasma:** Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store at -20°C. Avoid hemolysis and high cholesterol samples.
- **Other biological fluids:** Centrifuge at approximately 1000 × g for 20 min to remove precipitant. Analyze immediately or aliquot and store at -20°C or -80°C.

Note:

- » Fresh samples or recently obtained samples are recommended to prevent degradation and denaturalization that may lead to erroneous results. It is recommended to store samples to be used within 5 days at 4°C, within 1 month at -20°C and within 2 months at -80°C.
- » Samples should be clear and transparent. Samples must be diluted so that the expected concentration falls within the kit's range.
- » Please bring sample slowly to room temperature. Sample hemolysis will influence the result. Hemolyzed specimen should not be used. Samples that contain NaN₃ cannot be detected as it interferes with HRP.
- » Always use non-pyrogenic, endotoxin-free tubes for blood collection.

Sample dilution guideline:

Estimate the concentration of the target in the sample and select the correct dilution factor to make the diluted target concentration fall near the middle of the kit's range. Generally, for high concentration (40000 pg/ml - 400000 pg/ml), dilute 1:100, for medium concentration (4000 pg/ml - 40000 pg/ml), dilute 1:10 and for low concentration (62.5 pg/ml - 4000 pg/ml), dilute 1:2. Very low concentrations (≤ 62.5 pg/ml) do not need dilution. Dilute the sample with the provided Sample Diluent Buffer and mix thoroughly. Several trials may be necessary to determine the optimal dilution factor.

2. Wash buffer

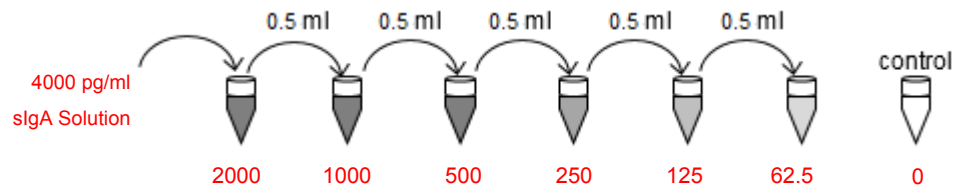
Dilute the concentrated Wash buffer 25-fold (1/25) with distilled water (i.e. add 30 ml of concentrated wash buffer into 720 ml of distilled water).

3. Standard

Preparation of the sIgA standard: standard solution should be prepared no more than 15 min prior to the experiment. Centrifuge at 10,000×g for 1 minute as the powder may drop off from the cap when opening. **(Note: Do not dilute the standard directly in the plate).** Once your standard has been reconstituted, it should be used right away. We do not recommend reusing the reconstituted standard.

a.) 4000 pg/ml standard solution. Add 1 ml of Sample/Standard diluent buffer into one Standard tube. Allow the reconstituted standard to sit for 15 minutes with gentle agitation prior to carrying out the serial dilutions; avoiding foaming or bubbles.

b.) 2000 pg/ml → 62.5 pg/ml standard solutions: Label 6 tubes with 2000 pg/ml, 1000 pg/ml, 500 pg/ml, 250 pg/ml, 125 pg/ml and 62.5 pg/ml. Aliquot 0.5 ml of the Sample / Standard diluent buffer into each tube. Add 0.5 ml of the above 4000 pg/ml standard solution into 1st tube and mix thoroughly. Transfer 0.5 ml from 1st tube to 2nd tube and mix thoroughly. Transfer 0.5 ml from 2nd tube to 3rd tube and mix thoroughly, and so on.



Note: Do not vortex the standard during reconstitution, as this will destabilize the protein. Once your standard has been reconstituted, it should be used right away. We do not recommend reusing the reconstituted standard. Please use the diluted Standards for a single assay procedure and discard after use.

4. Preparation of Biotin conjugated antibody working solution: prepare no more than 1 hour before the experiment.

- Calculate the total volume of the working solution: $0.1 \text{ ml / well} \times \text{quantity of wells}$. (Allow 0.1-0.2 ml more than the total volume).
- Dilute the Biotin conjugated antibody with antibody diluent buffer at 1/100 and mix thoroughly. i.e. Add 1 μl of Biotin conjugated antibody into 99 μl of antibody diluent buffer. Discard after use.

5. Preparation of HRP Conjugated Reagent working solution: prepare no more than 30 min. before the experiment

- Calculate the total volume of the working solution: $0.1 \text{ ml / well} \times \text{quantity of wells}$. (Allow 0.1-0.2 ml more than the total volume).
- Dilute the HRP Conjugate Reagent with HRP diluent buffer at 1/100 and mix thoroughly. i.e. Add 1 μl of HRP Conjugate Reagent into 99 μl of HRP diluent buffer. Discard after use.

5. Preparation of Substrate mixture solution:

- Calculate the total volume of the mixture solution: $0.1 \text{ ml / well} \times \text{quantity of wells}$. (Allow 0.1-0.2 ml more than the total volume).
- In a separate tube, add Substrate Reagent A and Substrate Reagent B in a 1:1 ratio and mix thoroughly. Discard after use.

B. Assay Procedure

- Set standard, test sample and control (zero) wells on the pre-coated plate and record their positions. It is recommended to measure each standard and sample in duplicate.
- Add 100 μl of the prepared standards solutions into the standard wells.
- Add 100 μl of Sample / Standard diluent buffer into the control (zero) well.
- Add 100 μl of appropriately diluted sample into test sample wells.
- Cover the plate and incubate at 37°C for 90 minutes.
- Remove the cover and discard the liquid. Do not wash.
- Add 100 μl of prepared Biotin conjugated antibody working solution into each well (standard, test sample and zero well). Add the solution at the bottom of each well without touching the side walls. Seal the plate with a cover and incubate at 37°C for 60 minutes.
- Remove the cover and discard the solution. Wash the plate 3 times with 1X Wash Buffer. Fill each well completely with Wash buffer (300 μL) using a multi-channel Pipette or autowasher (1-2 minute soaking period is recommended). Complete removal of liquid at each step is essential for good performance. After the final wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean absorbent paper towels.
- Add 100 μl of HRP working solution into each well, cover the plate and incubate at 37°C for 30 minutes.
- Remove the cover, discard the liquid and wash the plate 5 times with Wash Buffer as explained in step 8.

11. Add 100 µl of Substrate mixture solution into each well. Cover the plate and incubate at 37°C in dark conditions for up to 5 minutes.
12. Ensure that there are no fingerprints or water on the bottom of the plate, and that the fluid in the wells is free of bubbles. Measure the chemiluminescence signal in a microplate luminometer immediately.

For calculation, (the relative RLU) = (the RLU of each well) – (the RLU of Zero well). The standard curve can be plotted as the relative RLU of each standard solution (Y) vs. the respective concentration of the standard solution (X). Log-log curve fitting is recommended for data analysis. The slgA concentration of the samples can be interpolated from the standard curve.

Note: If the samples measured were diluted, multiply the dilution factor by the concentrations from interpolation to obtain the concentration before dilution.

C. Precautions

1. Equilibrate all reagents to room temperature (18-25°C) prior to use and centrifuge the tubes briefly in case any contents are trapped in the lid.
2. For each step in the procedure, total dispensing time for addition of reagents to the assay plate should not exceed 10 minutes.
3. Wash buffer may crystallize and separate. If this happens, please warm the tube and mix gently to dissolve.
4. Avoid foaming or bubbles when mixing or reconstituting components.
5. It is recommended to assay all standards, controls and samples in duplicate or triplicate.
6. Do NOT let the plate dry out completely during the assay as this will inactivate the biological material on the plate.
7. Ensure plates are properly sealed or covered during incubation steps.
8. Complete removal of all solutions and buffers during wash steps is necessary for accurate measurement readings.
9. To avoid cross contamination do not reuse pipette tips and tubes.
10. Do not use components from a different kit or expired ones.
11. The Substrate solutions are easily contaminated; work under sterile conditions when handling the Substrate solutions. The Substrate solutions should also be protected from light. Aspirate the dosage needed with sterilized tips and do not dump the residual solution back into the vial.

D. Precision

Intra-assay Precision (Precision within an assay): 3 samples with low, medium and high levels of slgA were tested 20 times on one plate, respectively.

Inter-assay Precision (Precision between assays): 3 samples with low, medium and high levels of slgA were tested on 3 different plates, 8 replicates in each plate.

$$CV (\%) = (\text{Standard Deviation} / \text{mean}) \times 100$$

Intra-Assay: CV<10%

Inter-Assay: CV<10%