

## Instructions for Use

Version: 1.0.3

Revision date: 23-Aug-24

# D-Galacturonic Acid Assay Kit

**Catalog No.:** abx298938

**Size:** 96 Assays

**Storage:** Store all components at 4°C.

**Application:** For quantitative detection of D-Galacturonic Acid concentrations in tissue homogenates, cell lysates, cell culture supernatants and other biological fluids.

**Detection Range:** 0.05 mmol/L - 2.5 mmol/L

**Introduction:** D-galacturonic acid is naturally occurring hexuronic acid that is a major component of plant cell wall polysaccharides and pectin. Abbexa's D-Galacturonic Acid Assay Kit is a quick, convenient, and sensitive method for measuring and calculating D-Galacturonic Acid concentrations. The dye reagents react with D-Galacturonic Acid to create an absorption maximum at 525 nm. The intensity of the color is proportional to the concentration of D-Galacturonic Acid, which can then be calculated.

### Kit components

1. 96 well microplate
2. Assay Buffer: 4 x 30 ml
3. Reaction Buffer: 15 ml
4. Dye Reagent: 1 vial
5. Dye Reagent Diluent: 1 ml
6. Standard: 1 vial
7. Plate sealer: 3

### Materials Required But Not Provided

1. Microplate reader (525 nm)
2. Distilled water
3. Pipettor, multi-channel pipettor
4. Pipette tips
5. Mortar
6. Centrifuge
7. Timer
8. Convection oven

## Protocol

### A. Preparation of Sample and Reagents

#### 1. Reagents

- **Standard Solution**

Add 1 ml of distilled water into the Standard vial and mix thoroughly. Ensure that the Standard has completely dissolved. Take 100  $\mu$ l of this solution and add 900  $\mu$ l of distilled water to prepare the Standard Solution (concentration 2.5 mmol/L).

- **Dye Reagent Solution**

Add 1 ml of Dye Reagent Diluent into the Dye Reagent vial and mix thoroughly to prepare the Dye Reagent Solution. Ensure that the Dye Reagent has completely dissolved prior to use.

#### 2. Sample

- **Cell and Bacterial samples**

Count the number of cells in the culture. Collect the cells into a centrifuge tube, and centrifuge to precipitate the cells. Discard the supernatant, and add 1 ml of Assay Buffer for every 5,000,000 cells. Sonicate at 20% power in 3 second bursts with a 10 second rest interval, for 30 bursts in total. Incubate at 90-95°C for 10 minutes, then centrifuge at 8000 x g at 4°C for 10 minutes. Transfer the supernatant to a new tube, keep on ice and analyze immediately.

- **Tissue samples**

Homogenize 0.1 g of sample in 1 ml of Assay Buffer, then incubate in a water bath at 80°C for 30 minutes. Centrifuge at 8000 x g at 4°C for 10 minutes. Transfer the supernatant to a new tube, keep on ice and analyze immediately.

- **Liquid samples**

Liquid samples can be used directly.

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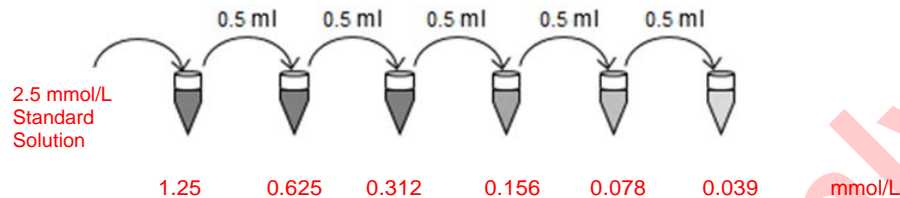
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### B. Assay Procedure

Bring all reagents to room temperature prior to use.

If the expected concentration is unknown, a preliminary experiment is recommended to determine if sample dilutions or adjustments to the reaction time are required to bring the measured concentration within the detection range of the kit.

- Label 6 tubes with 1.25 mmol/L, 0.625 mmol/L, 0.3125 mmol/L, 0.156 mmol/L, 0.078 mmol/L, and 0.039 mmol/L. Aliquot 0.5 ml of distilled water into each tube. Add 0.5 ml of 2.5 mmol/L Standard Solution to the 1<sup>st</sup> tube and mix thoroughly. Transfer 0.5 ml from the 1<sup>st</sup> tube to the 2<sup>nd</sup> tube and mix thoroughly, and so on.



- Set the sample, standard and blank wells on the 96 well microplate and record their positions. We recommend setting up each standard and sample in duplicate.
- Add 20  $\mu$ l of sample to the sample wells.
- Add 20  $\mu$ l of prepared standards to the standard wells.
- Add 20  $\mu$ l of distilled water to the blank wells.
- Add 150  $\mu$ l of Reaction Buffer to all wells.
- Tap the plate gently to mix. Cover the plate with a plate sealer, and incubate in a convection oven at 90°C for 20 minutes.
- Allow the plate to cool, then add 10  $\mu$ l of Dye Reagent Solution to all wells.
- Tap the plate gently to mix. Allow to stand for 2 minutes. Read and record absorbance at 525 nm.

### C. Calculations

D-galacturonic acid concentration per mg of protein:

$$\text{D-galacturonic acid } (\mu\text{mol/mg}) = \frac{C_{\text{Standard}} \times V_{\text{Standard}}}{V_{\text{Sample}} \times C_{\text{Protein}}} \times \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}} = \frac{2.5}{C_{\text{Protein}}} \times \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}}$$

D-galacturonic acid concentration per g of sample:

$$\text{D-galacturonic acid } (\mu\text{mol/g}) = \frac{C_{\text{Standard}} \times V_{\text{Standard}} \times V_{\text{Assay}}}{V_{\text{Sample}} \times W} \times \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}} = \frac{2.5}{W} \times \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}}$$

D-galacturonic acid concentration per 10<sup>4</sup> cells or bacteria:

$$\text{D-galacturonic acid } (\mu\text{mol}/10^4 \text{ cells}) = \frac{C_{\text{Standard}} \times V_{\text{Standard}} \times V_{\text{Assay}}}{V_{\text{Sample}} \times N} \times \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}} = \frac{2.5}{N} \times \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}}$$

D-galacturonic acid concentration per ml of sample:

$$\text{D-galacturonic acid } (\mu\text{mol/ml}) = \frac{C_{\text{Standard}} \times V_{\text{Standard}}}{V_{\text{Sample}}} \times \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}} = 2.5 \times \frac{\text{OD}_{\text{Sample}} - \text{OD}_{\text{Blank}}}{\text{OD}_{\text{Standard}} - \text{OD}_{\text{Blank}}}$$

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where:

<b>C<sub>Protein</sub></b>	Concentration of protein (mg/ml)
<b>C<sub>Standard</sub></b>	Concentration of highest standard (2.5 mmol/L= 2.5 $\mu$ mol/ml)
<b>W</b>	Weight of the sample (g)
<b>N</b>	Number of cells or bacteria ( $\times 10^4$ )
<b>V<sub>Assay</sub></b>	Volume of Assay Buffer (1 ml)
<b>V<sub>Sample</sub></b>	Volume of sample (0.02 ml)
<b>V<sub>Standard</sub></b>	Volume of standard (0.02 ml)

**D. Technical Support**For troubleshooting and technical assistance, please contact us at [support@abbexa.com](mailto:support@abbexa.com).

For Reference Only