

Human Coagulation Factor VIII (F8) CLIA Kit

Catalog No.: abx493207

Size: 96T

Range: 13.7 pg/ml - 10000 pg/ml

Sensitivity: < 4.9 pg/ml

Storage: Store standard, detection reagent A, detection reagent B and the 96-well plate at -20°C, and the rest of the kit components at 4°C.

Application: For quantitative detection of F8 in Human Plasma

Introduction: Factor VIII (FVIII) is a blood protein that promotes coagulation. It is typically bound to von Willebrand factor (vWF) to keep it inactive, but later separates from this protein in response to vascular damage. Active FVIII (FVIIIa) works as a cofactor to help factor IXa to cleave factor X, allowing the coagulation cascade to progress. FVIIIa is deactivated by factor IXa, protein C, and other proteases. Low levels of factor VIII result from the X-linked genetic disorder Haemophilia A, which leads to an increased bleeding time before coagulation and increases the risks from trauma. HIV-contaminated factor VIII products were responsible for a number of pharmaceutical scandals in the 1980s.

Principle of the Assay

This kit is based on chemiluminescent immunoassay technology. Anti-F8 antibody is pre-coated onto a 96-well plate. The standards and samples are added to the wells with a biotin conjugated anti-F8 and incubated. Next, Avidin conjugated to HRP is added to each microplate well and incubated. After the mixture of substrate A and B is added only wells that contain F8 will produce chemiluminescence. The intensity of the emitted light is proportional to the amount of F8 in the sample or standard.

Kit components

1. One pre-coated 96-well microplate (12 × 8 well strips)
2. Standard: 2 tubes
3. Standard Diluent Buffer: 20 ml
4. Wash Buffer (30X): 20 ml. Dilution: 1:30
5. Detection Reagent A (100X): 120 µl
6. Detection Reagent B (100X): 120 µl
7. Diluent A: 12 ml
8. Diluent B: 12 ml
9. Substrate A: 10 ml
10. Substrate B: 2 ml
11. Plate sealer: 4

Material Required But Not Provided

1. 37°C incubator
2. Luminometer capable of reading 96-well microplates (lag time 30.0 secs and read time 1.0 sec/well)
3. Multi and single channel pipettes and sterile pipette tips
4. Squirt bottle or automated microplate washer
5. Distilled or deionized water
6. Tubes to prepare standard or sample dilutions
7. Absorbent filter papers
8. 100 ml and 1 liter graduated cylinders

Protocol

A. Preparation of sample and reagents

1. Sample

Isolate the test samples soon after collecting, analyze immediately or store at 4°C for up to 5 days. Otherwise, store at -20°C for up to one month or -80°C for up to two months to avoid loss of bioactivity. Avoid multiple freeze-thaw cycles.

- **Plasma:** Collect plasma using 3.8% sodium citrate(sodium citrate : blood = 1: 9) as an anticoagulant. Centrifuge for 15 minutes at 1000 × g within 30 minutes of collection. Assay immediately or aliquot and store at -20°C or -80°C. Avoid hemolysis and high cholesterol samples.

Note:

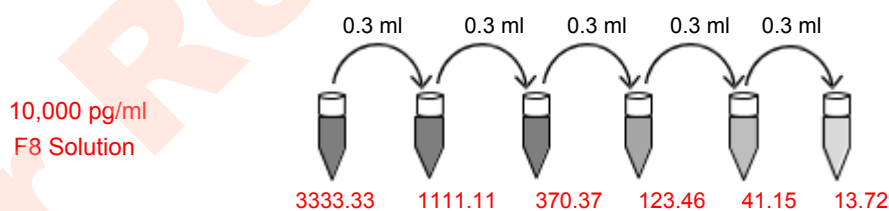
- » Bring sample slowly to room temperature. Sample hemolysis will influence the result. Hemolyzed specimen should not be used.
- » Samples must be diluted so that the expected concentration falls within the kit's range. Sample should be diluted in 0.01 mol/L PBS (PH=7.0-7.2). **Plasma samples require approximately a 100 fold dilution.**
- » If the sample are not indicated in the manual's applications, a preliminary experiment to determine the validity of the kit will be necessary.
- » Fresh sample or recently obtained samples are recommended to prevent protein degradation and denaturalization that may lead to erroneous results.
- » Always use non-pyrogenic, endotoxin-free tubes for blood collection.

2. Wash buffer

Dilute the concentrated Wash buffer 30-fold (1/30) with distilled water (i.e. add 20 ml of concentrated wash buffer into 580 ml of distilled water).

3. Standard

Bring samples and all kit components to room temperature. Prepare the Standard with 1.0 ml of Standard Diluent buffer (kept for 10 min at room temperature) to make the 10,000 pg/ml Standard Solution. Allow the reconstituted standard to sit for 10 minutes with gentle agitation prior to carrying out the serial dilutions; avoiding foaming or bubbles. Label 6 tubes with 3333.33 pg/ml, 1111.11 pg/ml, 370.37 pg/ml, 123.46 pg/ml, 41.15 pg/ml, 13.72 pg/ml. Aliquot 0.6 ml of the Standard diluent buffer into each tube. Add 0.3 ml of 10,000 pg/ml standard solution into the 1st tube and mix thoroughly. Transfer 0.3 ml from 1st tube to 2nd tube, mix thoroughly, and so on.



4. Detection Reagent A and B Preparation

Centrifuge Detection Reagent A and B briefly before use. Detection Reagent A and B should be diluted 100-fold with Diluent A and B respectively, and mixed thoroughly. They are sticky solutions, therefore pipette with a slow, smooth action to reduce volume errors. The solution should be prepared no more than 15 minutes prior to the experiment. Please discard after use.

5. Substrate working solution Preparation

Substrate A and B should be mixed with a ration of 99:1 respectively and mix thoroughly. For example, prepare 1 ml of substrate working solution by mixing 0.99 ml of Substrate A and 0.01 ml of Substrate B.

B. Assay Procedure

Equilibrate the kit components and samples to room temperature before use. It is recommended to plot a standard curve for each test.

1. Set standard, test sample and control (zero) wells on the pre-coated plate respectively, and then, record their positions. It is recommended to measure each standard and sample in duplicate. Add the solution at the bottom of each well without touching the side walls. Mix the standards and samples up and down to be homogeneous before adding into the wells but avoid adding bubbles.
2. Add 100 µl of the diluted standards into the standard wells. Aliquot 100 µl Standard Diluent Buffer to the control (zero) well.
3. Add 100 µl of appropriately diluted sample into the test sample wells. Add the solution at the bottom of each well without touching the side wall. Shake the plate mildly to mix thoroughly.
4. Seal the plate with a cover and incubate for 1 h at 37°C.
5. Remove the cover and discard the liquid. Do not wash.
6. Aliquot 100 µl of the detection Reagent A working solution to each well. Seal the plate with a cover and incubate for 1 h at 37°C.
7. Remove the cover and discard the solution. Wash the plate 3 times with 1X Wash Buffer. Fill each well completely with Wash buffer (350 µl) using a multi-channel Pipette or autowasher (1-2 minute soaking period is recommended). Complete removal of liquid at each step is essential for good performance. After the final wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean absorbent paper towels.
8. Aliquot 100 µl of Detection Reagent B working solution into each well, seal and incubate at 37°C for 30 min.
9. Discard the solution and wash the plate 5 times with wash buffer as explained in step 7 (each time let the Wash Buffer stay for 1-2 min).
10. Aliquot 100 µl of Substrate working solution into each well. Seal the plate with a cover and incubate at 37°C for 10 min. Avoid exposure to light. The incubation time is for reference only, the optimal time should be determined by the end user.
11. Measure the chemiluminescence signal in a microplate luminometer immediately.

For calculation, average the duplicate readings for each set of reference standard, control and samples and subtract the average zero standard RLU (Relative Light Unit). The standard curve can be plotted with the mean RLU of each reference standard on the Y axis vs. the respective concentration of each standard solution on the X axis. The F8 concentration of the samples can be interpolated from the standard curve.

Note: If the samples measured were diluted, multiply the dilution factor by the interpolated concentration of the sample to obtain the concentration before dilution.

C. Precautions

1. Before using the kit, centrifuge the tubes briefly to bring down the contents trapped in the lid.
2. If crystals have formed in the concentrated Wash Buffer, warm to room temperature and mix gently until the crystals have completely dissolved.
3. Avoid foaming or bubbles when mixing or reconstituting components. Prepare the Standard dilutions within 15 min of use and discard any unused working standards. For each step in the procedure, total dispensing time for addition of reagents to the assay plate should not exceed 10 minutes.
4. It is recommended measuring each standard and sample in duplicate.
5. Do not let the wells uncovered for extended periods between incubation. Once reagents are added to the wells, avoid letting the strips dry as this can inactivate the biological material on the plate. Incubation time and temperature must be controlled.
6. Ensure plates are properly sealed or covered during incubation steps.
7. Complete removal of all solutions and buffers during wash steps is necessary for accurate measurement readings.
8. Do not reuse pipette tips and tubes to avoid cross contamination.
9. Do not vortex the standard during reconstitution, as this will destabilize the protein. Once your standard has been reconstituted, it should be used right away. We do not recommend reusing the reconstituted standard.
10. The Substrate solution is easily contaminated; work under sterile conditions when handling the substrate solution. The Substrate A and B should also be protected from light. Unreacted substrate should be colorless or very light yellow in appearance. Aspirate the dosage needed with sterilized tips and do not dump the residual solution back into the vial.

D. Precision

Intra-assay Precision (Precision within an assay): 3 samples with low, medium and high levels of F8 were tested 20 times on one plate, respectively.

Inter-assay Precision (Precision between assays): 3 samples with low, medium and high levels of F8 were tested on 3 different plates, 8 replicates in each plate.

$$CV (\%) = (\text{Standard Deviation} / \text{mean}) \times 100$$

Intra-Assay: CV<10%

Inter-Assay: CV<12%