

# Instructions for Use

Version: 1.0.0

Revision date: 25 Sep 2020



## Human Kruppel Like Factor 15 (KLF15) CLIA Kit

**Catalog No:** abx494237

**Size:** 96T

**Range:** 0.156 ng/ml - 10 ng/ml

**Sensitivity:** < 0.057 ng/ml

**Storage:** Store the 96-well plate, Standards, and Detection Reagent(s) at -20°C, and the rest of the kit components at 4°C.

**Application:** The quantitative detection of KLF15 in Human tissue homogenates, cell lysates, cell culture supernatant and other biological fluids.

**Principle of the Assay:** This kit is based on sandwich chemiluminescent immunoassay technology. An antibody is pre-coated onto a 96-well plate. Standards, test samples, and biotin-conjugated reagent are added to the wells and incubated. The HRP-conjugated reagent is then added, and the whole plate is incubated. Unbound conjugates are removed using wash buffer at each stage. After the Substrate working solution is added, only wells that contain the labelled KLF15 will produce chemiluminescence. The intensity of the emitted light is proportional to the amount of KLF15 in the sample.

### Kit Components

- Pre-coated 96-Well Microplate: 12 x 8
- Standard: 2 tubes
- Standard Diluent Buffer: 20 ml
- Wash Buffer: (30X) 20 ml
- Detection Reagent A: (100X) 120 µl
- Detection Reagent B: (100X) 120 µl
- Diluent A: 12 ml
- Diluent B: 12 ml
- Substrate A: 10 ml
- Substrate B: 2 ml
- Plate Sealer: 4

### Materials Required But Not Provided

- 37°C incubator
- Multi and single channel pipettes and sterile pipette tips
- Squirt bottle or automated microplate washer
- 1.5 ml tubes
- Distilled water
- Absorbent filter papers
- 100 ml and 1 liter graduated cylinders
- 0.01 mol/L PBS (pH 7.0 - 7.2)
- Luminometer capable of reading 96-well microplates (lag time 30.0 secs and read time 1.0 sec/well)

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## Protocol

### A. Sample Preparation

Analyse immediately or store samples at 2-8°C (within 24 hrs). For long term storage, aliquot and store at -20°C or -80°C. Avoid multiple freeze-thaw cycles.

- **Tissue Homogenates:** The preparation of tissue homogenates will vary depending upon tissue type – this is just an example. Rinse tissues with ice-cold PBS to remove the excess of blood. Weigh before homogenization. Finely mince tissues and homogenize with a tissue homogenizer on ice in PBS and sonicate the cell suspension. Centrifuge the homogenates at 5000 × g for 5 mins and collect the supernatant.
- **Cell Lysates:** Detach adherent cells with trypsin and collect by centrifugation and remove the supernatant. Wash the cells three times in ice-cold PBS and re-suspend cells in PBS. Lyse the cells by ultra-sonification 4 times, or freeze at -20°C and thaw to room temperature 3 times. Centrifuge at 1500 × g for 10 mins at 2-8°C to remove cellular debris. Collect the supernatant.
- **Cell Culture Supernatant:** Centrifuge at approximately 1000 × g for 20 mins to remove precipitate.
- **Other Biological Fluids:** Centrifuge at approximately 1000 × g for 20 mins to remove precipitate. Analyse immediately or aliquot and store at -20°C or -80°C.

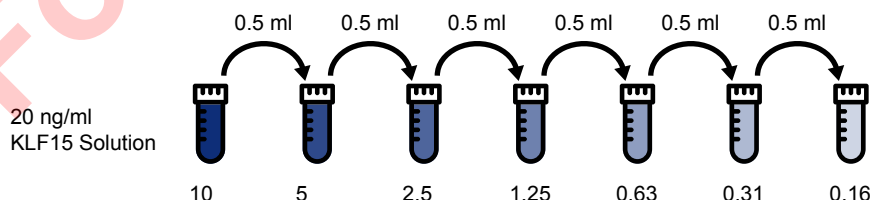
### Notes:

- Samples must be diluted so that the expected concentration falls within the kit's range. The sample should be diluted in 0.01 mol/L PBS (PH=7.0-7.2).
- Fresh samples, or recently obtained samples, are recommended to prevent protein degradation and denaturation that may lead to erroneous results.
- NaN<sub>3</sub> cannot be used as a test sample preservative, since it inhibits HRP.
- If a sample is not indicated in the manuals applications, a preliminary experiment to determine the suitability of the kit will be required.

### B. Reagent Preparation

**Standard:** Prepare the Standard with 1 ml of Standard Diluent buffer to make the 20 ng/ml Standard Solution. Allow the reconstituted standard to sit for 10 mins, with gentle agitation prior to carrying out the serial dilutions. Avoid foaming or bubbles. Further dilute by a factor of 2 to give the highest standard, 10 ng/ml. Label tubes in preparation for the serial dilutions - *see the diagram below for reference*. Aliquot 0.5 ml of the Standard Diluent Buffer into each tube (apart from the highest standard tube). Add 0.5 ml of the highest standard solution into the 1st tube and mix thoroughly. Transfer 0.5 ml from the 1st to 2nd tube, mix thoroughly, and so on.

**Note:** Do not vortex the standard during reconstitution, as this will destabilize the protein. Once the standard has been reconstituted, it should be used within 15 mins. It is not recommended to reuse the reconstituted standard.



**Wash Buffer:** Dilute the concentrated Wash Buffer 30-fold (1/30) with distilled water (i.e. add 20 ml of concentrated wash buffer into 580 ml of distilled water). If crystals have formed in the concentrated Wash Buffer, warm to room temperature and mix gently until the crystals have completely dissolved.

**Detection Reagent A Working Solution Preparation:** Prepare no more than 15 mins before the experiment.

1. Calculate the total volume of working solution required.
2. Dilute Detection Reagent A 100-fold with Diluent A, and mix thoroughly. Pipette with a slow, smooth action to reduce volume errors.

**Detection Reagent B Working Solution Preparation:** Prepare no more than 15 mins before the experiment.

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1. Calculate the total volume of working solution required.
2. Dilute Detection Reagent B 100-fold with Diluent B, and mix thoroughly. Pipette with a slow, smooth action to reduce volume errors.

## **Preparation of Substrate Working Solution:**

1. Calculate the total volume of working solution required.
2. In a separate tube, add Substrate A and Substrate B in a 99:1 ratio, respectively (e.g. add 990  $\mu$ l TMB Substrate A to 10  $\mu$ l TMB Substrate B, respectively. Prepare no more than 10 mins before use.

## **C. Assay Protocol**

Prepare all standards, samples and reagents as directed above. Equilibrate the kit components and samples to room temperature prior to use. It is recommended to measure in duplicate, and to plot a standard curve for each test.

1. Set standard, test sample and control (zero) wells on the pre-coated plate respectively, and record their positions. *Add the solution to the bottom of each well without touching the side walls. Pipette the standards and samples up and down to mix before adding to the wells. Avoid foaming or bubbles.*
2. Aliquot 100  $\mu$ l of the diluted standards into the standard wells.
3. Aliquot 100  $\mu$ l of Standard Diluent buffer into the control (zero) well.
4. Aliquot 100  $\mu$ l of appropriately diluted sample into the test sample wells. Gently tap the plate to mix, or use a microplate shaker.
5. Cover the plate with a plate sealer and incubate for 1 hr at 37°C.
6. Remove the cover and discard the liquid. Do not wash.
7. Aliquot 100  $\mu$ l of Detection Reagent A working solution to each well. Cover the plate with a plate sealer and incubate for 1 hr at 37°C.
8. Remove the cover and discard the solution. Wash the plate 3 times with Wash Buffer. *Fill each well completely with Wash buffer (300  $\mu$ l) using a multi-channel Pipette or autowasher (1-2 mins soaking period is recommended). Complete removal of liquid at each step is essential for good performance. After the final wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean absorbent paper towels.*
9. Aliquot 100  $\mu$ l of Detection Reagent B working solution to each well. Seal the plate and incubate for 30 mins at 37°C.
10. Remove the cover, discard the solution and repeat the wash process described in Step 8, 5 times.
11. Aliquot 100  $\mu$ l of Substrate working solution into each well. Seal the plate with a cover and incubate at 37°C for 5-10 mins. The incubation time is for reference only, and the optimal time should be determined by end user.
12. Ensure that there are no fingerprints or water on the bottom of the plate, and that the fluid in the wells is free of bubbles. Measure the chemiluminescence signal in a microplate luminometer immediately.

For calculation, average the duplicate readings for each set of reference standard, control and samples and subtract the average zero standard RLU (Relative Light Unit). The standard curve can be plotted with the mean RLU of each reference standard on the Y axis vs. the respective concentration of each standard solution on the X axis. The KLF15 concentration of the samples can be interpolated from the standard curve. **Note:** If the samples measured were diluted, multiply the dilution factor by the interpolated concentration of the sample to obtain the concentration before dilution.

## **Precautions:**

- Before using the kit, centrifuge the tubes to bring down the contents trapped in the lid.
- Do not leave the wells uncovered for extended periods between incubations. The addition of reagents for each step should not exceed 10 mins.
- Ensure that the plate is properly sealed or covered during the incubation steps, and that the time and temperature are controlled.
- Do not reuse pipette tips and tubes.
- Do not use expired components, or components from a different kit.
- The TMB substrate should be used under sterile conditions, and light exposure should be minimised. Unused substrate should be colorless, or a very light yellow in appearance. Do not discard any residual solution back into the vial.

## **Precision:**

Intra-assay Precision (Precision within an assay): 3 samples with low, medium and high levels of KLF15 were tested 20 times on one plate,

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respectively.

Inter-assay Precision (Precision between assays): 3 samples with low, medium and high levels of KLF15 were tested on 3 different plates, 8 replicates in each plate.

$CV (\%) = (\text{Standard Deviation} / \text{Mean}) \times 100$

Intra-Assay:  $CV < 10\%$

Inter-Assay:  $CV < 12\%$

For Reference Only