ABL1 (pY245) Cell ELISA Kit

Catalog No.: abx596013

Size: 96T

Range: Qualitative

Sensitivity: Qualitative

Storage: Store at 4°C for up to 6 months.

Application: For qualitative detection of ABL1 (pY245) in cell samples.

Principle of the Assay

This kit is based on indirect enzyme-linked immuno-sorbent assay technology. Cells are seeded on to a 96-well plate. The cells are fixed and quenched, and the wells are blocked. The primary antibody specific to ABL1, ABL1 (pY245), or GAPDH antibody is added to each well, followed by incubation. Fluorescent dye-conjugated secondary antibody is used as the detection antibody, which is added to each well, followed by incubation. Unbound conjugates are washed away using wash buffer. The RFU of each well is measured at excitation/emission wavelengths 651/667 nm (Dye 1) or at 495/521 nm (Dye 2).

Kit components

1. 96-well black cell culture microplate: 2
2. TBS (10X): 24 ml
3. Quenching buffer: 24 ml
4. Blocking buffer: 50 ml
5. GAPDH mouse monoclonal primary antibody (100X): 110 µl
6. ABL1 (pY245) primary antibody (100X): 60 µl
7. ABL1 primary antibody (100X): 60 µl
8. Dye 1-conjugated Anti-Rabbit IgG Antibody: 6 ml
9. Dye 2-conjugated Anti-Mouse IgG Antibody: 6 ml
10. Primary antibody diluent buffer: 12 ml
11. Wash buffer (15X): 50 ml
12. Plate Sealer: 2

Material Required But Not Provided

1. 37°C incubator
2. Fluorescent plate reader (two channels at Ex/Em: 651/667 & 495/521 nm)
3. Multi and single channel pipettes and sterile pipette tips
4. Squirt bottle or automated microplate washer
5. ELISA orbital shaker
6. Deionised or distilled water
7. Fixing solution (4%, 8%, or 37% paraformaldehyde)
8. Poly-L-Lysine (if using suspension cells)
9. Tissue culture grade water
10. 1.5 ml tubes to prepare standard/sample dilutions
11. Absorbent filter papers
12. 100 ml and 1 liter graduated cylinders
13. Parafilm

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Protocol

A. Experimentatal design and Preparation of sample and reagents

1. Experimental design and Sample preparation

- The number of cells plated on to the 96-well plate will depend on the level of expression of ABL1 (pY245) in the cells, cell size, treatment conditions and incubation time.
- Cells can be treated with inhibitors, activators, stimulators (e.g. chemicals, proteins, peptides) or a combination of these. Stimulation of cells should be controlled.
- Positive control: Mouse Anti-GAPDH Antibody is used as an internal positive control to normalize the RFU values of the target protein in each well.
- Negative control: Add 50 µl of primary antibody diluent buffer instead of the primary antibody mixture solution. Add the 50 µl secondary antibody mixture solution as described in the assay procedure (i.e. only the secondary antibody mixture solution is incubated).
- Blank control: Add 50 µl of primary antibody diluent buffer instead of the primary antibody mixture solution or the secondary antibody mixture solution (i.e. no antibody mixture solution is incubated).

2. Wash buffer and TBS

- Dilute the concentrated Wash buffer 15-fold (1/15) with distilled water (i.e. add 10 ml of concentrated wash buffer into 140 ml of distilled water).
- Dilute the concentrated TBS 10-fold (1/10) with distilled water (i.e. add 10 ml of concentrated TBS into 90 ml of distilled water).

3. Preparation of primary antibody mixture solution (Phospho): Prepare the mixture no more than 15 min. before the experiment. Add 50 µl of 100X anti-ABL1 (pY245) antibody and 50 µl of 100X GAPDH antibody into 4900 µl of Primary antibody diluent buffer, and mix thoroughly. Label as "Primary Antibody Mixture Phospho".

4. Preparation of primary antibody mixture solution (Non-phospho): Prepare the mixture no more than 15 min. before the experiment. Add 50 µl of 100X anti-ABL1 antibody and 50 µl of 100X GAPDH antibody into 4900 µl of Primary antibody diluent buffer, and mix thoroughly. Label as "Primary Antibody Mixture Non-Phospho".

5. Preparation of secondary antibody mixture solution: prepare no more than 15 min. before the experiment.
For each plate, add 3 ml of Dye 1-conjugated Anti-Rabbit IgG Antibody and 3 ml of Dye 2-conjugated Anti-Mouse IgG Antibody together, and mix thoroughly. Label as "Secondary Antibody Mixcture" This solution is light sensitive and should be kept in the dark.

B. Assay Procedure

1. Seed 100 µl of 20,000 adherent cells in each well of a sterile 96-well plate. Add solutions slowly from the side wall of the wells. The optimal seeding concentration should be determined by the end user.
   For suspension cells and loosely attached cells: Add 20 µl of Poly-L-Lysine to each well of the 96-well plate and allow to stand for 5 minutes. Remove the solution by aspiration and rinse once with tissue culture grade water. Allow to dry for 2 hours before seeding cells. If the cell line is known to digest Poly-L-Lysine, Poly-D-Lysine may be used as an alternative.
2. Incubate the cells for at least 6 hours (or overnight) at 37°C, 5% CO_2.
3. Treat the cells as desired. Please consider the treatment method as cell treatment may result in cell death.
4. Remove the cell culture medium gently. Fix the cells by adding 100 µl of fixing solution and incubating for 20 minutes at room temperature. A solution of 4% paraformaldehyde is recommended for adherent cells and 8% paraformaldehyde is recommended for suspension cells and loosely attached cells. If 37% paraformaldehyde is used, dilute to the desired percentage using 1X PBS.
5. Remove the Fixing solution and wash the plate 3 times with 1X Wash buffer. Fill each well completely with 1X Wash buffer (300 µl) using a multi-channel pipette or autowasher. A 3 minute soaking period with gentle shaking on an orbital shaker is recommended. After the final wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and tap **gently** against clean absorbent paper towels. Complete removal of liquid at each step is essential for good performance.

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6. Add 100 µl of Quenching buffer into each well and incubate for 20 minutes at room temperature.
7. Wash the plate 3 times as directed in Step 5.
8. Add 200 µl of Blocking buffer into each well and incubate for 1 hour at room temperature.
9. Designate Phospho and Non-Phospho wells, and add 50 µl of each primary antibody mixture solution to the appropriate wells for detection. Designate 2 wells as negative controls, and do not add primary antibody mixture solutions to these wells. Cover the plate and incubate overnight at 4°C. If the target expression is known to be high, incubate for 2 hours at room temperature with gentle shaking on an orbital shaker.
10. Wash the plate 3 times as directed in Step 5.
11. Add 50 µl of secondary antibody mixture solution to the appropriate wells. Cover the plate and incubate for 1.5 hours at room temperature with gentle shaking on an orbital shaker.
12. Wash the plate 3 times as directed in Step 5.
13. Read the plate at Ex/Em 651/667 nm (Dye 1) and 495/521 nm (Dye 2). Avoid direct light exposure.

C. Precautions

1. Before using the kit, centrifuge the tubes briefly to bring down any contents trapped in the lid.
2. For each step in the procedure, total dispensing time for addition of reagents to the assay plate should not exceed 10 minutes.
3. The wash buffer may crystallise and separate. If this happens, please warm the tube and mix gently to dissolve.
4. Avoid foaming or bubbles when mixing or reconstituting components.
5. It is recommended to assay all controls and samples in duplicate or triplicate.
6. Complete removal of all solutions and buffers during wash steps is necessary for accurate measurement readings.
7. To avoid cross contamination do not reuse pipette tips and tubes.
8. Do not use expired components or components from a different kit.
9. The Fixing solution is volatile and the Stop solution is corrosive. Wear Personal Protective Equipment (PPE) such as lab coats, goggles, mask and gloves when handling these solutions.
10. Vigorous pipetting my cause cells to detatch from the plate. Pipette solutions and aspirate gently.

D. Data Normalisation

Intra-well GAPDH normalisation: The measured RFU primary antibody values can be normalised using the RFU GAPDH antibody values using (RFU primary antibody (Em. 667) / RFU GAPDH antibody (Em. 521)).

To analyze the effects of stimulants, the non-phospho primary antibody allows normalisation between phosphorylated- and non-phosphorylated values.

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