

Mouse IL-6 Receptor Subunit Beta / GP13 (IL6ST) ELISA Development Kit

Catalogue No.:abx378347

Mouse IL-6 Receptor Subunit Beta / GP13 (IL6ST) ELISA Development Kit for use in Sandwich ELISA assay development.

This ELISA Development Kit contains:

Component	5 × 96 tests	15 × 96 tests
Pretreated 96-well ELISA Plate	5	15
Capture Antibody	120 μΙ	350 μΙ
Biotin-Conjugated Detection Antibody	120 μΙ	350 μΙ
HRP-Conjugate	120 µl	350 μΙ
Standard (200 ng)	1 vial	15 vials

Please note that quantities and concentrations may change between different batches.

It is recommended to use this ELISA Development Kit with <u>abx471002 ELISA Development Support Kit (Sandwich Method)</u>; alternatively, the following solutions can be prepared separately:

Coating Buffer: 1X Citrate-Buffered Saline

Blocking Buffer: 1X PBS containing 0.5-3% BSA

• Wash Buffer: 3% Tris

Standard and Sample Diluent Buffer: 1X PBS containing 0.5-3% BSA

• Antibody and HRP-Conjugate Diluent Buffer: 1X PBS containing 0.5-3% BSA

• Stop Solution: 5% Sulfuric Acid

Target: IL-6 Receptor Subunit Beta / GP13 (IL6ST)

Reactivity: Mouse

Tested Applications: ELISA

Recommended dilutions: Capture Antibody: 1/500 - 1/1000, Biotin-conjugated Detection Antibody: 1/500 - 1/1000,

HRP-Conjugate: 1/500 - 1/1000. Optimal dilutions/concentrations should be determined by

the end user.

Reconstitution: Reconstitute the standard with 1 ml of Standard Diluent to obtain a stock standard solution

of 200 ng/ml. Label tubes in preparation for the serial dilutions (100, 50, 25, 12.5, 6.25, and 3.125 ng/ml). Aliquot 0.5 ml of the Standard Diluent into each tube. Add 0.5 ml of the highest standard solution into the 1st tube and mix thoroughly. Transfer 0.5 ml from the 1st to 2nd

tube, mix thoroughly, and so on.

Storage: Aliquot and store at -20°C in the dark. Avoid repeated freeze/thaw cycles.

Datasheet

Version: 1.0.0 Revision date: 29 Mar 2025



UniProt Primary AC: Q00560 (UniProt, ExPASy)

Test Range: 3.12 ng/ml - 200 ng/ml

Detection Method: Colormetric

Assay Type: Sandwich

Sample Type: Serum and plasma.

Sample Collection/Preparation: Analyse immediately or store samples at 2-8°C (within 24 hrs). For long term storage,

aliquot and store at -20°C or -80°C. Avoid multiple freeze-thaw cycles.

• **Serum:** Samples should be collected into a serum separator tube. Coagulate the serum by leaving the tube undisturbed in a vertical position overnight at 4°C or at room temperature for up to 1 hr. Centrifuge at approximately 1000 × g for 20 mins. If precipitate appears, centrifuge again. Assay immediately or aliquot and store at -20°C or -80°C.

• Plasma: Collect plasma using EDTA as an anticoagulant. Centrifuge for 15 mins at 1000 × g, within 30 mins of collection. If precipitate appears, centrifuge again. Avoid hemolytic samples.

Note: THIS PRODUCT IS FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC,

THERAPEUTIC OR COSMETIC PROCEDURES. NOT FOR HUMAN OR ANIMAL

CONSUMPTION.



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Directions for use:

Bring all components to room temperature (18-25°C) and briefly spin or centrifuge the vials before use. Working solutions should be prepared and used immediately.

Recommended Procedure:

- 1. Dilute the Capture Antibody to working concentration using Coating Buffer. Immediately coat the 96-well plate with diluted Capture Antibody (100 µl per well). Seal the plate and incubate at 2-8 °C overnight.
- 2. Remove the liquid from each well. Do not wash. Block the plate with Blocking Buffer (200 μ l per well) at 37 °C for 1 hour.
- 3. Remove the liquid from each well. Do not wash. Either proceed with the following steps immediately or dry the plate at 37 °C for 30 minutes, then store at -20 °C with dessicant for up to 6 months.
- 4. Add 100 μ l of standards or sample into the appropriate wells. Cover with a plate sealer and incubate at 37 °C for 1.5 hours.
- 5. Remove the liquid from each well. Do not wash. Add appropriately diluted Biotin-Conjugated Detection Antibody (100 μ l per well). Cover the plate with a new plate sealer and incubate at 37 °C for 1 hour.
- 6. Remove the liquid from each well. Wash with Wash Buffer (350 μl per well) and allow to soak for 1-2 min. Remove the liquid by inverting and tapping the plate on to absorbent paper. Repeat the wash process 3 times.
- 7. Add appropriately diluted HRP-Conjugate (100 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 30 min.
- 8. Repeat the wash process in Step 6, for a total of 5 times.
- 9. Add Substrate Solution (90 µl per well). Cover the plate with a new plate sealer and incubate at 37 °C for 15-30 min. Keep the plate in the dark and avoid exposure to light.
- 10. Add Stop Solution (50 µl per well). Tap the side of the plate to ensure thorough mixing.
- 11. Measure the absorbance immediately using a microplate reader set at 450 nm.